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## Increasing muscle co-contraction speeds up internal model acquisition during dynamic motor learning

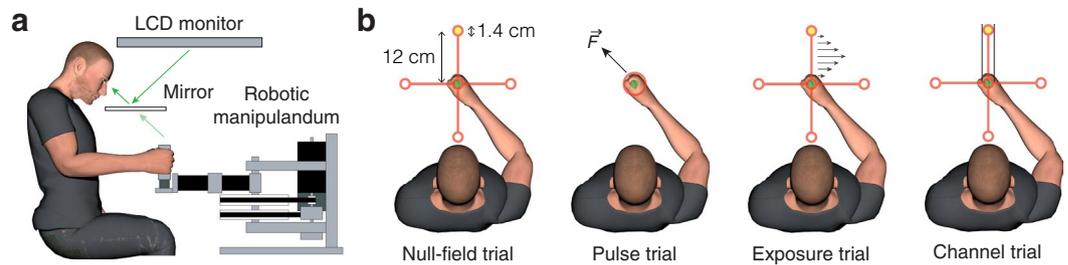
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During reaching movements in the presence of novel dynamics, participants initially co-contract their muscles to reduce kinematic errors and improve task performance. As learning proceeds, muscle co-contraction decreases as an accurate internal model develops. The initial co-contraction could affect the learning of the internal model in several ways. By ensuring the limb remains close to the target state, co-contraction could speed up learning. Conversely, by reducing kinematic errors, a key training signal, it could slow down learning. Alternatively, given that the effects of muscle co-contraction on kinematic errors are predictable and could be discounted when assessing the internal model error, it could have no effect on learning. Using a sequence of force pulses, we pretrained two groups to either co-contract (stiff group) or relax (relaxed group) their arm muscles in the presence of dynamic perturbations. A third group (control group) was not pretrained. All groups performed reaching movements in a velocity-dependent curl field. We measured adaptation using channel trials and found greater adaptation in the stiff group during early learning. We also found a positive correlation between muscle co-contraction, as measured by surface electromyography, and adaptation. These results show that muscle co-contraction accelerates the rate of dynamic motor learning.

The reduction of kinematic errors during reaching movements in the presence of novel dynamics occurs through at least two complementary mechanisms<sup>1,2</sup>. First, participants can learn an internal model of the dynamics and use this model to anticipate and compensate for perturbations<sup>3–6</sup>. Second, participants can use a less-specific strategy of stiffening up the limb through muscle co-contraction (i.e., impedance control)<sup>1,7–9</sup> to reduce the kinematic errors that result from perturbing dynamics<sup>10–12</sup>. Several studies have shown that both these mechanisms contribute to the early compensation for novel dynamics<sup>13–17</sup> and that muscle co-contraction decreases as the internal model is learned<sup>1,18–20</sup>. Therefore, muscle co-contraction offers a temporary, non-specific strategy to compensate for novel dynamics until they are learned by the internal model. Here we ask whether the increase in muscle co-contraction during exposure to novel dynamics affects the learning of the internal model.

Muscle co-contraction could affect dynamic motor learning in three contrasting ways. First, it could facilitate learning by keeping the state of the limb close to the desired state required for task success (i.e., the target). By resisting force-field displacements, muscle co-contraction increases the alignment between the actual motions experienced and the motion to be learned. Importantly, because participants learn the mapping from visited states, rather than planned states, to dynamics<sup>21</sup>, increased muscle co-contraction should accelerate learning by concentrating adaptation within the region of state space associated with the planned motion. Indeed, in artificial neural networks, it has been shown that the inverse dynamics of a plant can be learned faster when the actual trajectories executed are closer to the desired trajectories to be learned, as this ensures that local error information is applicable to the region of state space to be learned<sup>22</sup>. Conversely, reduced muscle co-contraction should slow down learning by increasing the range of actual motions experienced and thus spreading adaptation more

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**Figure 1.** Paradigm. (a) The participant grasped the handle of the robotic manipulandum (vBOT) while seated. Visual feedback was presented veridically using a horizontally-mounted monitor viewed through a mirror. (b) At the start of each reaching trial, a target was illuminated (yellow circle), cueing the participant to move their hand (green circle) from the center of the workspace to the illuminated target by making a fast reaching movement. In the pulse phase, a series of brief pulses of force (black arrow) was applied to the handle of the manipulandum in a random direction. Participants in the stiff group were instructed to keep their hand within a 1.5 cm radius of the home position (red ring) by co-contracting the muscles of their arm. Participants in the relaxed group were instructed to let their hand move freely by relaxing the muscles of their arm. In the exposure phase, a velocity-dependent force field was applied. Channel trials were used to assess adaptation for the 90° target only.

thinly across state space. Consistent with this idea, adaptation to a visuomotor rotation is less complete when the rotation is introduced abruptly compared with gradually<sup>23</sup>.

Second, muscle co-contraction could impede learning by reducing kinematic variability<sup>10,24–26</sup>, which has been shown in some forms to enhance error-based learning, both in humans<sup>27</sup> and songbirds<sup>28–30</sup>. Moreover, muscle co-contraction reduces kinematic errors, which are a key training signal for internal model acquisition<sup>31–34</sup>. Although some saturation is seen for large errors, in general, trial-to-trial learning increases with the magnitude of the error on the previous trial<sup>35,36</sup>. Therefore, by reducing kinematic errors, muscle co-contraction could reduce trial-to-trial learning and thus decrease the overall rate or extent of learning.

Third, muscle co-contraction could have no effect on learning if kinematic errors are normalized against the current level of muscle co-contraction. Indeed, incoming sensory information is often pre-processed on the basis of motor output. For example, reafferent feedback is attenuated by sensory predictions based on efference copies of motor commands<sup>37–39</sup>. It is therefore possible that error-based learning is driven by pre-processed kinematic errors. Consequently, even though kinematic errors are smaller during muscle co-contraction, the motor system may still adapt to the true internal model performance error. Alternatively, or concurrently, the opposing effects of concentrated learning in state space (hypothesis one) and smaller kinematic errors (hypothesis two) could cancel out such that muscle co-contraction has no effect on learning.

To differentiate between these hypotheses, we developed a novel paradigm in which participants were either pretrained to increase (stiff group) or decrease (relaxed group) muscle co-contraction in the presence of dynamic perturbations. A third group (control group) was not pretrained at all. We then examined force-field adaptation using channel trials, which constrain movements to a straight line and allow us to measure the force generated into the channel walls as a measure of predictive compensation. In the initial stage of exposure, the stiff group adapted more than both the relaxed group and the control group. In both the initial and final stage of exposure, we found a positive correlation between an individual's level of muscle co-contraction and their level of adaptation.

## Materials and Methods

Thirty-six neurologically intact participants (16 men and 20 women; age  $25.3 \pm 5.3$  yr, mean  $\pm$  s.d.) were recruited to participate in the experiment, which had been approved by the Cambridge Psychology Research Ethics Committee and was performed in accordance with guidelines and regulations. All participants provided written informed consent and were right-handed according to the Edinburgh handedness inventory<sup>40</sup>. Participants had not previously performed any experiments involving velocity-dependent curl fields.

**Apparatus.** Experiments were performed using a vBOT planar robotic manipulandum<sup>41</sup> with virtual-reality system and air table (Fig. 1a). The vBOT is a modular, general-purpose, two-dimensional planar manipulandum optimized for dynamic learning paradigms. Position was measured using optical encoders sampled at 1 kHz. Torque motors allowed forces to be generated at the endpoint. A monitor mounted above the vBOT projected visual feedback into the plane of movement via a horizontal mirror. The location of the mirror prevented direct vision of the hand and forearm. Participants were seated in front of the robotic manipulandum with their torso restrained by a four-point harness to reduce movement. The handle of the manipulandum was grasped with the right hand, while the right forearm was supported on an air sled, which constrained arm movements to the horizontal plane.

Movements were performed in either a null field (vBOT passive), a velocity-dependent curl force field or in a simulated channel. In the curl field, the force generated at the handle of the manipulandum was given by:

$$\begin{bmatrix} f_x \\ f_y \end{bmatrix} = b \begin{bmatrix} 0 & -1 \\ 1 & 0 \end{bmatrix} \begin{bmatrix} \dot{x} \\ \dot{y} \end{bmatrix} \quad (1)$$

where  $f_x$ ,  $f_y$ ,  $\dot{x}$  and  $\dot{y}$  are the forces and velocities at the handle in the  $x$ - and  $y$ -directions respectively. The gain  $b$  was set to  $\pm 16$  Ns/m, where the sign of  $b$  specified the direction of the curl (clockwise or counterclockwise). On channel trials, the hand was constrained to move along a straight line from the home position to the target, using stiffness (6,000 N/m) and damping (5 Ns/m) perpendicular to the movement direction. Channel trials clamped the kinematic error to zero, allowing forces that were generated in a feedforward manner to be measured orthogonal to the direction of reach<sup>13,42</sup>.

**Electromyography.** Surface electromyography (EMG) was recorded from two monoarticular shoulder muscles (pectoralis major and posterior deltoid), two biarticular shoulder-elbow muscles (biceps brachii and the long head of the triceps) and two monoarticular elbow muscles (the lateral head of the triceps and brachioradialis). The EMG was recorded using the Delsys Bagnoli 8 electromyography system (Boston, MA). The skin was cleaned with alcohol and prepared with an abrasive gel to increase conductance. After the abrasive gel was removed with cotton wool, conductive gel was applied to the electrodes, which were secured to the skin with tape. Electrode placement was chosen to maximize signal while minimizing crosstalk from adjacent muscles. EMG signals were analog band-pass filtered between 20 and 450 Hz (in the Delsys Bagnoli EMG system) and then sampled at 2 kHz.

**Paradigm.** With the exception of the pulse phase, in which a series of force pulses was applied to the handle of the robot while the hand was stationary, participants made 12 cm center-out reaching movements to one of four targets located at 0°, 90°, 180° and 270°. Visual feedback consisted of the home position (0.7 cm radius disk), the four targets (0.7 cm radius rings), a set of red lines connecting each target to the home position and the hand cursor (0.5 cm radius disk) (Fig. 1b). The four targets, the home position and the connecting lines were displayed at all times, except during the pulse phase. Each trial started when the hand cursor had remained within 0.3 cm of the home position at a speed below 0.5 cm/s for 100 ms. One of the targets was then filled in, followed 500 ms later by an auditory tone, which cued the participant to initiate their movement. The trial ended when the hand cursor had remained within 0.3 cm of the target position for 500 ms. If the peak speed of the movement was less than 40 cm/s or greater than 60 cm/s, a low-frequency auditory tone was played, and a ‘Too Slow’ or ‘Too Fast’ message was displayed, respectively. At the end of each trial, the hand was passively returned to the home position by the robotic manipulandum.

Participants were assigned to either a stiff group ( $n = 12$ ), a relaxed group ( $n = 12$ ) or a control group ( $n = 12$ ). Each experiment began with a training phase to familiarize participants with the passive dynamics of the robot and the requirements of the task. Participants then performed a pre-exposure phase consisting of 100 null-field trials (20 blocks), a pulse phase consisting of 50 trials (stiff and relaxed groups only), a post-pulse phase consisting of four null-field trials, an exposure phase consisting of 410 force-field trials (83 blocks) and a post-exposure phase consisting of 100 null-field trials (20 blocks). Each block of five trials consisted of one null or field trial to each target and one channel trial to the 90° target. We only applied channel trials for one of the four targets (the 90° target for all participants to allow comparison) to provide enough force-field trials for learning without making the experiment excessively long. The order of trials within a block was pseudorandom, except that a channel trial never occurred on the first trial of a block. The force field direction was fixed for each participant but counterbalanced across participants.

The pulse phase was designed to increase muscle co-contraction in the stiff group and decrease muscle co-contraction in the relaxed group. Each trial started with the hand in the home position. A single force pulse (5 N for 500 ms) was then applied to the hand. The 50 force pulses (one for each trial) ranged uniformly in direction over 360° and were presented in a pseudorandom order. Participants in the stiff group were instructed to keep the handle positioned within a border (1.5 cm radius ring) around the home position by co-contracting the muscles of their limb (Fig. 1b). Participants in the relaxed group were instructed to let their hand move freely by relaxing the muscles of their limb. After the force pulse had ended, the handle of the manipulandum was passively returned to the home position in preparation for the next trial.

During the exposure phase, participants in the stiff group were instructed to maintain their arm stiff and to not deviate from the red lines connecting the home position to the target (described above), and participants in the relaxed group were instructed to relax their arm. The control group did not perform the pulse phase and were not given any explicit instructions regarding muscle co-contraction. The exact instructions given to participants can be found in the Supplementary Information.

Participants were given a 45 second rest break after the pre-exposure phase and after every 13 blocks (65 trials) in the exposure phase, with the exception of the end of the exposure phase, which was followed directly by the post-exposure phase. At the end of each rest break, participants in the stiff group were reminded that co-contracting their muscles may help them to make straight reaching movements, and participants in the relaxed group were reminded that if they relaxed their arm their movements would naturally become straighter. To mitigate the effects of time-dependent decay of memory, a channel trial did not occur in the first four movements after each rest break.

**Analysis.** Data analysis was performed using MATLAB R2017b. Two measures of performance were computed. On exposure and null-field trials the maximum distance between the path of the hand and a straight line connecting the initial hand position and the target (maximum perpendicular error) was computed. On channel trials the percentage of the force field that was compensated for (adaptation) was computed by regressing the actual forces  $f_a(t)$  generated by participants in the channel on the ideal forces  $f_i(t)$  that would fully compensate for the forces on a force-field trial (defined in Eq. 1):

$$f_a(t) = k \times f_i(t), \quad (2)$$

$$\text{adaptation} = k \times 100\%, \quad (3)$$

where  $k$  is the regression coefficient and  $t$  is the discrete time step of the channel trial. The offset of the regression was constrained to zero. For this analysis, we used the portion of the movement where the hand speed was greater than 1 cm/s. At all stages of the experiment the ideal force term  $f_i(t)$  refers to the force required to fully compensate for the force field present in the exposure phase. Therefore, adaptation in the pre-exposure phase prior to learning should be close to zero. To combine maximum perpendicular error (MPE) and adaptation data across participants, data from individual participants was sign adjusted according to the direction of the field they were trained on.

To quantify oscillations on individual channel trials, we calculated an oscillation index that was the path length of each force profile (from when the hand speed exceeded 1 cm/s to when it dropped below 1 cm/s), where a greater path length indicates a more oscillatory profile. To control for differences in movement duration and adaptation level, each force profile was first normalized in amplitude by dividing by the force range (max - min) and normalized in time between 0 and 1.

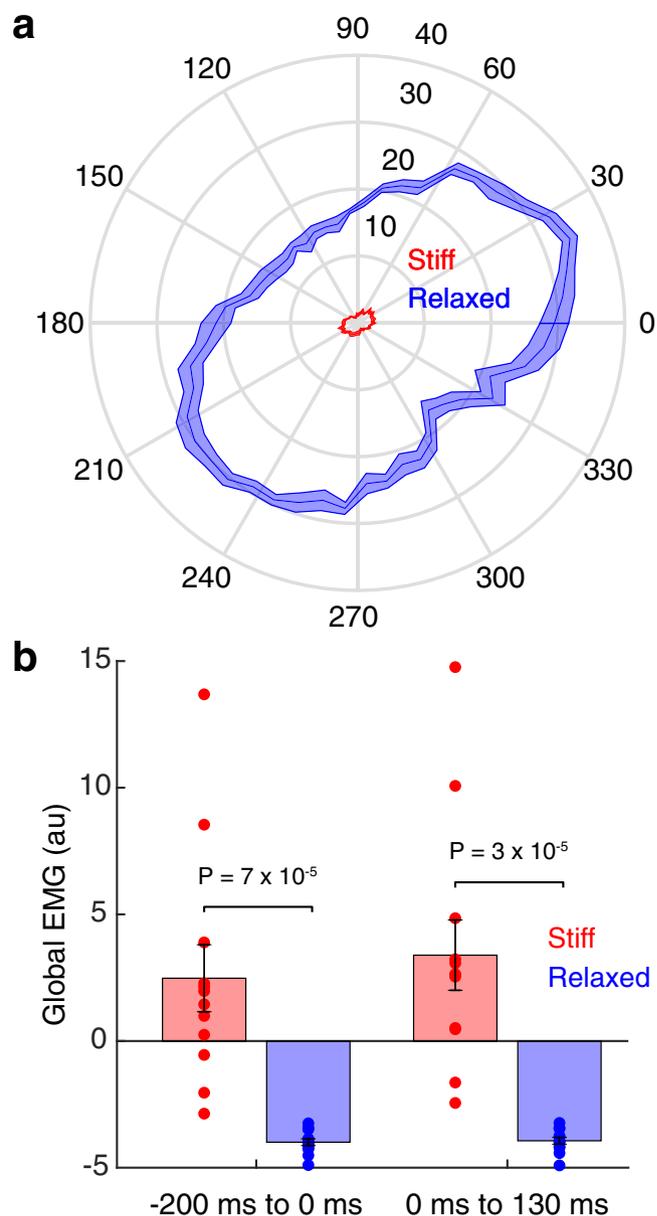
The raw EMG signal was bandpass filtered (30–500 Hz) using a 10<sup>th</sup>-order Butterworth filter implemented with MATLAB's `filtfilt` function and then full-wave rectified. We examined the EMG over two periods: an early period comprised of feedforward signals and a later period comprised of both feedforward and feedback signals. These periods were defined separately for the pulse phase and for reaching movements. In the pulse phase, the early period was taken from 200 ms before the perturbation onset to the perturbation onset, and the later period was taken from the perturbation onset to 130 ms after the perturbation onset. During reaching movements, the early period was taken from 200 ms before movement onset to 130 ms after movement onset. This period can be used as a measure of the feedforward muscle activation as feedback components of EMG are first detectable around 130 ms after movement onset when the hand is smoothly perturbed by a velocity-dependent force field<sup>16</sup>. The later period was taken from 130 ms to 400 ms after movement onset. To normalize EMG traces across the muscles, we first computed for each muscle the mean EMG over the early period for the non-channel trials in the pre-exposure phase. We then normalized the EMG trace for each trial by dividing the trace by the corresponding mean for that muscle<sup>19</sup>. Note that this only scales the EMG traces for each muscle and does not affect the temporal profile. A global measure of muscle activity was then computed for each trial by calculating the difference between the summed EMG across muscles (first averaged across time points on a trial) and the summed pre-exposure EMG across muscles (first averaged across time points and trials for the pre-exposure phase). Accordingly, negative global EMG signifies reduced activity relative to the pre-exposure phase, whereas positive global EMG signifies increased activity relative to the pre-exposure phase. For plotting purposes, EMG waveforms were time normalized by resampling 1000 data points between movement onset and movement end on each trial for each participant. Movement onset was defined as the time the cursor first moved 0.3 cm from the center of the home position. Movement end was defined as the time the cursor was first within 0.3 cm of the center of the target position. Each resampled data point was assigned a time point linearly spaced between 0 and the mean movement duration across participants.

To identify differences in global EMG, MPE and adaptation between the groups, between-subjects ANOVAs, mixed-design ANOVAs and unpaired  $t$ -tests were performed. We also performed linear regression between global EMG, adaptation and oscillation index. All statistical tests were two-sided with significance set to  $P < 0.05$ . Data are reported as mean  $\pm$  standard error of the mean (s.e.m.).

## Results

Participants started the experiment by making center-out reaching movements in a null field (pre-exposure phase). Then, in the stiff and relaxed groups, voluntary changes to muscle co-contraction were elicited by instructing participants to resist or comply with a sequence of force pulses while the hand was stationary (pulse phase). Figure 2a shows the maximum distance moved by the hand following force pulses in different directions. The maximum distance moved by the hand was smaller in the stiff group compared with the relaxed group (stiff: 1.7 cm  $\pm$  0.1, relaxed: 23.2  $\pm$  0.9, two-tailed unpaired  $t$ -test,  $t_{22} = 22.81$ ,  $P = 8 \times 10^{-17}$ ). This was due to greater muscle co-contraction, as confirmed by global EMG (stiff: 2.9  $\pm$  1.4, relaxed:  $-4.0 \pm 0.1$ , mixed-design ANOVA,  $F_{1,22} = 25.74$ ,  $P = 4 \times 10^{-5}$ ), both in the period preceding the force pulse (Fig. 2b; stiff: 2.5  $\pm$  1.3, relaxed:  $-4.0 \pm 0.1$ , two-tailed unpaired  $t$ -test,  $t_{22} = 4.87$ ,  $P = 7 \times 10^{-5}$ ) and in the period following pulse onset (Fig. 2b; stiff: 3.4  $\pm$  1.4, relaxed:  $-3.9 \pm 0.1$ , two-tailed unpaired  $t$ -test,  $t_{22} = 5.26$ ,  $P = 3 \times 10^{-5}$ ). The variability of global EMG was greater for the stiff group (Fig. 2b). This is most likely because the relaxed group are bounded by an EMG of zero, whereas the stiff group can stiffen up to different extents due to different bounds across participants. Note that because joint impedance increases faster than force variability as a function of muscle co-contraction<sup>24</sup>, co-contraction has a net stabilizing effect on endpoint kinematics<sup>25,43</sup>. The pulse phase was omitted for the control group.

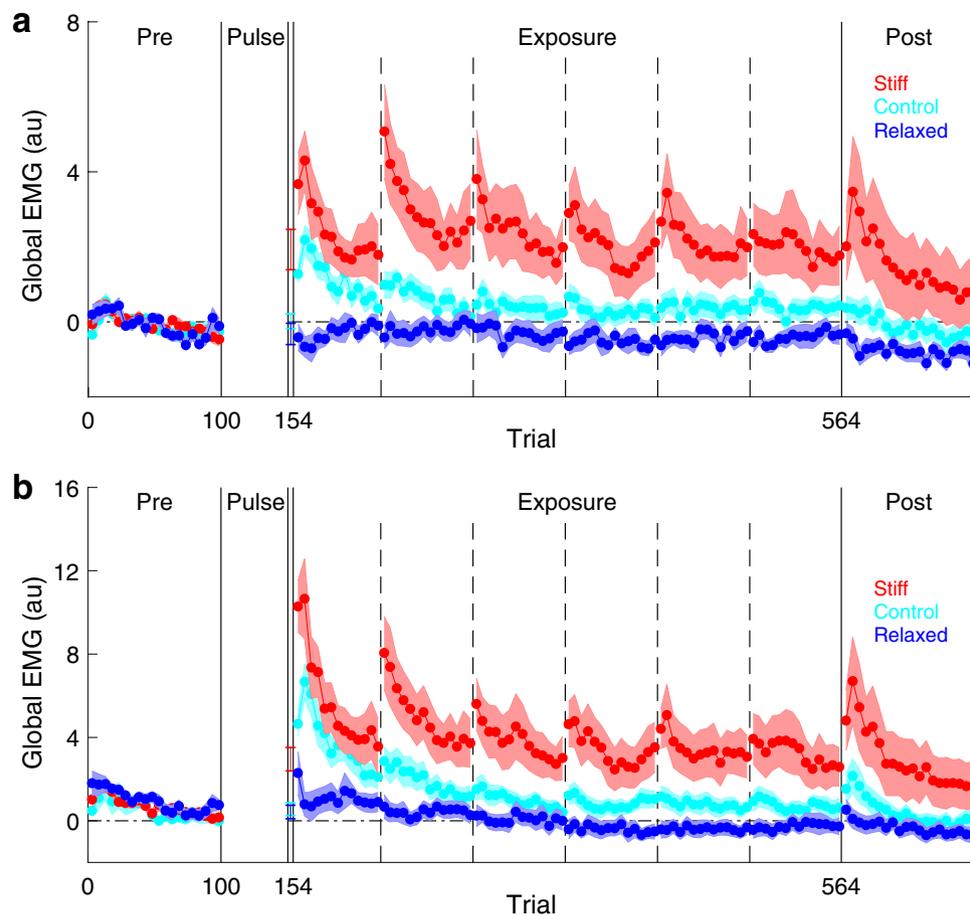
**Electromyography.** After the pulse phase (stiff and relaxed groups) or the pre-exposure phase (control group), participants performed reaching movements in a velocity-dependent curl field (exposure phase). Figure 3 shows global EMG for all groups over the course of the experiment, split into periods separated by rest breaks (single-participant data shown in Supplementary Figs 1 and 2). To identify differences in muscle co-contraction between the groups during the exposure phase, we performed a mixed-design ANOVA with stage of exposure (first vs. second half) and period of movement (early vs. later period) as within-subject factors and global EMG as the response variable. We found a main effect of group (mixed-design ANOVA,  $F_{2,33} = 8.59$ ,  $P = 0.001$ ) but no



**Figure 2.** Performance in the force pulse phase. **(a)** Polar plot of the maximum distance (cm) the hand moved as a function of the force pulse direction. Data show mean  $\pm$  s.e.m. across participants. **(b)** Global EMG before (left) and after (right) the force pulse for the stiff (red) and relaxed (blue) groups. Data show mean  $\pm$  s.e.m. across participants.

interaction between group and stage of exposure (mixed-design ANOVA,  $F_{2,33} = 1.58$ ,  $P = 0.220$ ). We therefore combined data from the first and second halves of the exposure phase. Importantly, global EMG was greater in the stiff group than the relaxed group, both in the early period of the movement (Fig. 3a; stiff:  $2.3 \pm 0.8$ , relaxed:  $-0.4 \pm 0.2$ , two-tailed unpaired  $t$ -test,  $t_{22} = 3.42$ ,  $P = 0.002$ ) and in the later period of the movement (Fig. 3b; stiff:  $4.1 \pm 1.1$ , relaxed:  $0.1 \pm 0.3$ , two-tailed unpaired  $t$ -test,  $t_{22} = 3.44$ ,  $P = 0.002$ ). Global EMG was also greater in the stiff group than the control group, both in the early period of the movement (Fig. 3a; control:  $0.5 \pm 0.2$ , two-tailed unpaired  $t$ -test,  $t_{22} = 2.28$ ,  $P = 0.033$ ) and in the later period of the movement (Fig. 3b; control:  $1.5 \pm 0.4$ , two-tailed unpaired  $t$ -test,  $t_{22} = 2.20$ ,  $P = 0.038$ ). Finally, Global EMG was greater in the stiff group than the relaxed group, both in the early period of the movement (Fig. 3a; two-tailed unpaired  $t$ -test,  $t_{22} = 3.15$ ,  $P = 0.005$ ) and in the later period of the movement (Fig. 3b; two-tailed unpaired  $t$ -test,  $t_{22} = 2.70$ ,  $P = 0.013$ ). Therefore, as expected, the stiff group exhibited the greatest degree of muscle co-contraction, followed by the control group and then the relaxed group.

Figure 4 shows the average EMG of the stiff (red), relaxed (blue) and control (cyan) groups during the first half of the exposure phase. This shows an increase in EMG both prior to and during movement in many of the muscles. Overall, a clear voluntary increase in EMG can be seen for the stiff group compared with the relaxed group.



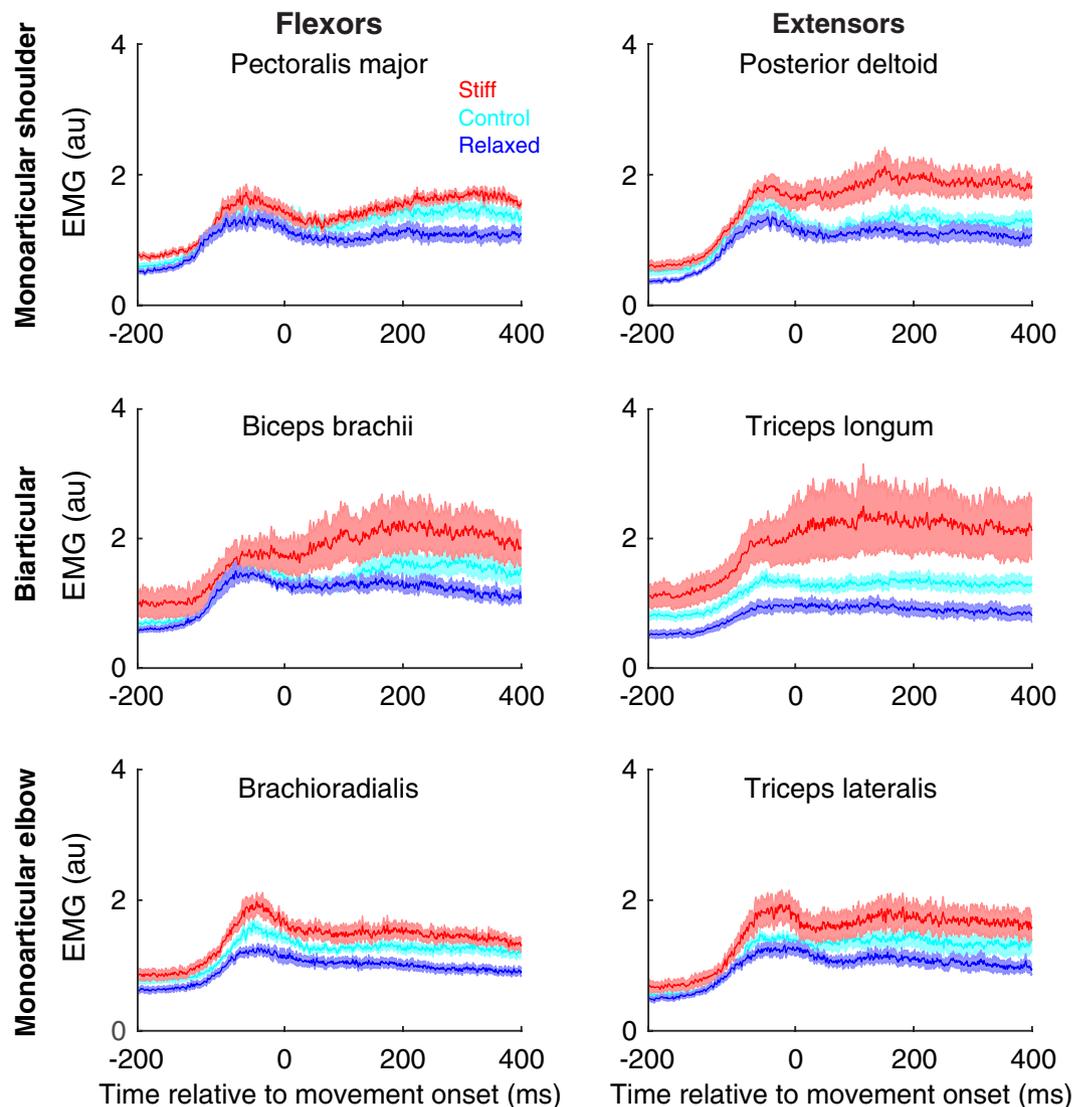
**Figure 3.** Global EMG over the course of the experiment. **(a,b)** Global EMG, which is a proxy for stiffness, was calculated on each trial and plotted as an average of a block of five trials. Data show mean  $\pm$  s.e.m. across participants in the stiff (red), relaxed (blue) and control (cyan) groups. Vertical dashed lines indicate rest breaks. **(a)** Global EMG from  $-200$  ms to  $130$  ms relative to movement onset. **(b)** Global EMG from  $130$  ms to  $400$  ms relative to movement onset.

This increase is seen in all muscles pairs, suggesting that muscle co-contraction is not specific to a single joint. The EMG of the control group is intermediate between the stiff and relaxed groups.

**Movement Analysis.** In the pre-exposure phase, reaching movements were relatively straight in all groups, as demonstrated by small MPEs (Fig. 5a; single-participant data shown in Supplementary Fig. 3). During the exposure phase, there was no difference in peak velocity between the groups (stiff:  $48.8 \pm 0.4$  cm/s, relaxed:  $48.4 \pm 0.5$ , control:  $48.0 \pm 0.5$ , between-subjects ANOVA,  $F_{2,33} = 0.61$ ,  $P = 0.549$ ). Therefore, each group experienced similar forces during learning. In the first block (5 trials) of the exposure phase, all groups showed large deviations from a straight line, and these deviations were different between the groups (between-subjects ANOVA,  $F_{2,33} = 9.94$ ,  $P = 4 \times 10^{-4}$ ). Importantly, MPE was, as expected, smaller in the stiff group compared with the relaxed group (stiff:  $3.3$  cm  $\pm$  0.2, relaxed:  $4.5 \pm 0.2$ , two-tailed unpaired  $t$ -test,  $t_{22} = 5.42$ ,  $P = 2 \times 10^{-5}$ ). Furthermore, MPE was also smaller in the control group compared with the relaxed group (control:  $3.2 \pm 0.3$ , two-tailed unpaired  $t$ -test,  $t_{22} = 3.49$ ,  $P = 0.002$ ). However, there was no difference in MPE between the stiff group and the control group (two-tailed unpaired  $t$ -test,  $t_{22} = 0.03$ ,  $P = 0.977$ ). As the exposure phase progressed, MPE decreased in all groups. By the final 13 blocks (65 trials) of the exposure phase, there was no difference in MPE between the groups (stiff:  $0.4$  cm  $\pm$  0.1, relaxed:  $0.7 \pm 0.1$ , control:  $0.7 \pm 0.2$ , between-subjects ANOVA,  $F_{2,33} = 1.94$ ,  $P = 0.160$ ). In the null field of the post-exposure phase, MPE rose again, demonstrating after-effects.

In the relaxed group, increases in MPE can be seen at the start of each rest break (Fig. 5a; vertical dashed lines) when participants were reminded that their movements would naturally become straighter if they relaxed their arm. In the stiff group, changes in MPE cannot be seen at the start of each rest break. This is likely because decreases in MPE due to increased co-contraction are offset by increases in MPE due to forgetting of learning during rest breaks.

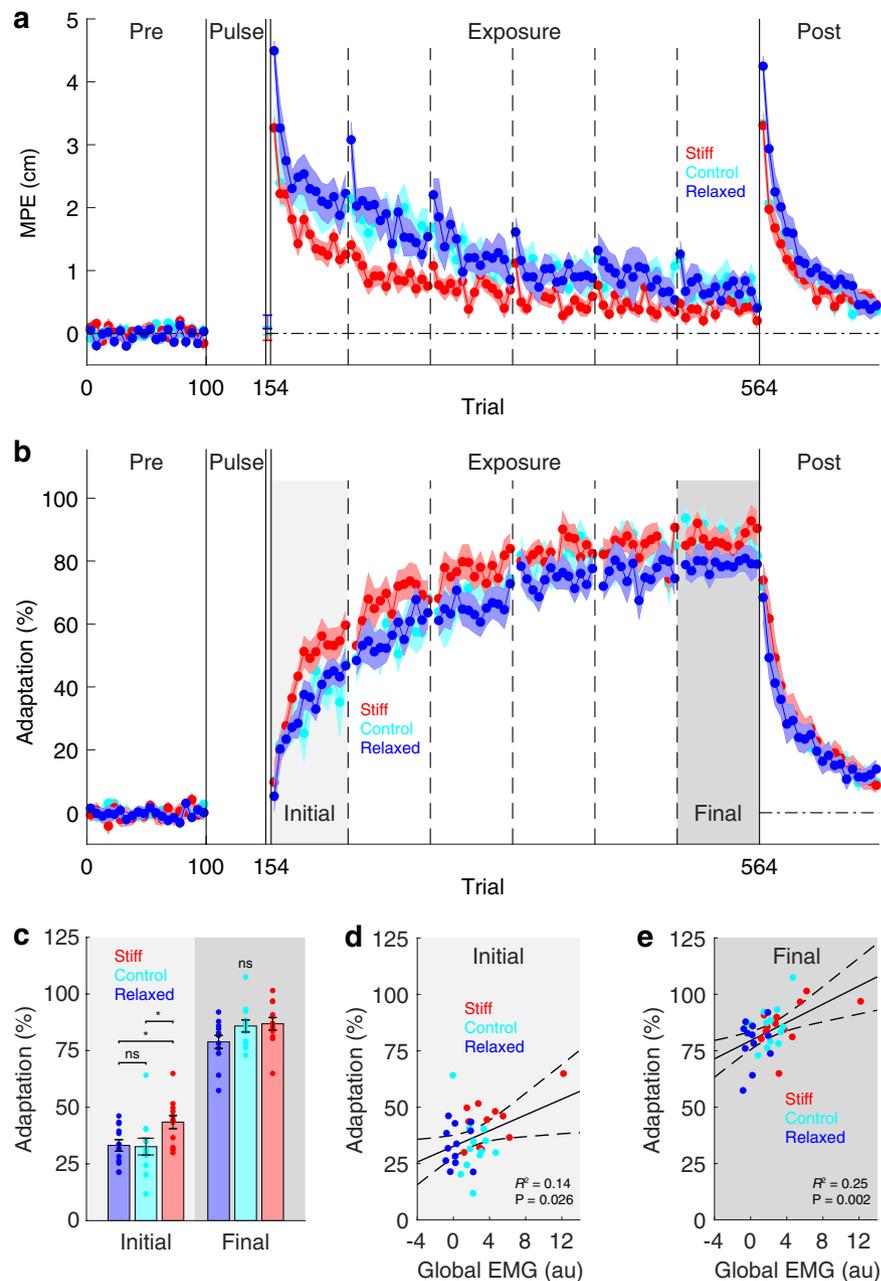
**Force compensation.** In the pre-exposure phase, adaptation measured on channel trials was, as expected, close to zero in all groups (Fig. 5b; single-participant data shown in Supplementary Fig. 4). During exposure, adaptation increased in all groups, eventually reaching asymptote. To assess learning, we compared initial (first 13



**Figure 4.** Muscle activity during the first half of the exposure phase. The EMG is averaged across all directions on force field trials. The period after movement onset has been time normalized. The time from 0 ms to 400 ms corresponds to approximately 64% of the mean movement duration across participants. The period before movement onset is shown in real time. Data show mean  $\pm$  s.e.m. across all participants in the stiff (red), relaxed (blue) and control (cyan) groups.

blocks of the exposure phase) and final (last 13 blocks of the exposure phase) adaptation between the groups. This revealed a main effect of group (mixed-design ANOVA,  $F_{2,33} = 3.75$ ,  $P = 0.034$ ) and no interaction (mixed-design ANOVA,  $F_{2,33} = 2.35$ ,  $P = 0.111$ ). Initial adaptation was greater in the stiff group than both the relaxed group (Fig. 5c; stiff:  $43.5 \pm 2.9$ , relaxed:  $33.2 \pm 2.5$ , two-tailed unpaired  $t$ -test,  $t_{22} = 2.68$ ,  $P = 0.014$ ) and the control group (Fig. 5c; control:  $32.6 \pm 3.7$ , two-tailed unpaired  $t$ -test,  $t_{22} = 2.31$ ,  $P = 0.031$ ). However, there was no difference in initial adaptation between the relaxed group and the control group (Fig. 5c; two-tailed unpaired  $t$ -test,  $t_{22} = 0.12$ ,  $P = 0.904$ ). Final adaptation was not significantly different between the groups (Fig. 5c; stiff:  $86.9 \pm 2.8$ , relaxed:  $78.9 \pm 2.9$ , control:  $85.9 \pm 2.7$ , between-subjects ANOVA,  $F_{2,33} = 2.49$ ,  $P = 0.099$ ).

We examined data at the individual level pooling across all three groups. This revealed a significant linear relationship between a participant's global EMG (first 13 blocks of the exposure phase, early and later periods of the movement combined) and both their initial adaptation (Fig. 5d;  $R^2 = 0.14$ ,  $\beta = 1.74$ ,  $F$ -test,  $F_{1,34} = 5.45$ ,  $P = 0.026$ ) and their final adaptation (Fig. 5e;  $R^2 = 0.25$ ,  $\beta = 2.02$ ,  $F$ -test,  $F_{1,34} = 11.17$ ,  $P = 0.002$ ). To rule out the possibility that muscle co-contraction influences adaptation by increasing/decreasing oscillations on channel trials, we performed multiple linear regression with global EMG and oscillation index (see methods) as predictors and adaptation as the response variable. Importantly, oscillation index was not a significant predictor of either initial adaptation ( $\beta = -0.17$ , two-tailed  $t$ -test,  $t_{33} = 0.04$ ,  $P = 0.971$ ) or final adaptation ( $\beta = -7.58$ , two-tailed  $t$ -test,  $t_{33} = 1.00$ ,  $P = 0.327$ ). In contrast, global EMG was a significant predictor of both initial adaptation ( $\beta = 1.75$ , two-tailed  $t$ -test,  $t_{33} = 2.19$ ,  $P = 0.036$ ) and final adaptation ( $\beta = 2.25$ , two-tailed  $t$ -test,  $t_{33} = 3.24$ ,  $P = 0.003$ ). Therefore, the relationship between global EMG and adaptation cannot be explained in terms of increased/decreased oscillations on channel trials due to muscle co-contraction.



**Figure 5.** Kinematic and dynamic adaptation over the course of the experiment. **(a)** Maximum perpendicular error (MPE) was calculated on each trial and plotted as an average of a block of five trials. Data show mean  $\pm$  s.e.m. across participants in the stiff (red), relaxed (blue) and control (cyan) groups. Vertical dashed lines indicate rest breaks. **(b)** Is the same as **(a)** but for adaptation measured on channel trials. **(c)** Mean  $\pm$  s.e.m. initial adaptation (first 13 blocks of the exposure phase) and final adaptation (last 13 blocks of the exposure phase) for all groups. Asterisks indicate significant ( $*P < 0.05$ ), ns indicates not significant. **(d,e)** Least-squares regression between each participant's global EMG (first 13 blocks of the exposure phase, early and later periods of movement combined) and their initial adaptation **(d)** and their final adaptation **(e)**. Dashed curves show 95% confidence intervals for the fitted linear model. Data from all groups have been pooled.

## Discussion

We examined force-field adaptation under increased, decreased and baseline levels of muscle co-contraction. Participants were randomly assigned to either a stiff, relaxed or control group. The stiff and relaxed groups were pretrained using a sequence of force pulses to either increase (stiff group) or decrease (relaxed group) muscle co-contraction in the presence of perturbations. The control group was not pretrained. All groups performed reaching movements in a velocity-dependent curl field. We assessed learning by measuring adaptation, that is predictive force compensation using channels trials. In the initial stage of exposure, the stiff group adapted more than the relaxed and control groups, despite experiencing smaller kinematic errors, an important training signal for error-based learning. Furthermore, at the individual level, muscle co-contraction was significantly correlated

with adaptation, both in the initial and final stage of exposure. These results show that, in addition to improving kinematic accuracy, muscle co-contraction also increases the rate of acquisition of an internal model.

There are a number of reasons why increased muscle co-contraction could lead to an increase in the rate of dynamic motor learning. First, when motor errors are experienced, the internal model is updated with respect to the actual states visited rather than the intended states, a process termed motion-referenced learning<sup>21</sup>. Consequently, any intervention that increases the overlap between the actual motions experienced and the motion required to reach the target, such as increased muscle co-contraction, could increase the rate of adaptation. Second, error sensitivity is greater for smaller errors<sup>34,36,44</sup>. This could explain why muscle co-contraction accelerates adaptation, despite decreasing the size of errors. In a causal inference framework<sup>45</sup>, larger errors are more likely to have been caused by factors extrinsic to the motor plant and so should be discounted to a greater extent. Consequently, larger errors do not guarantee greater learning. Indeed, single-trial adaptation (the product of error sensitivity and error magnitude) appears to be non-monotonic with a peak at an intermediate error magnitude<sup>46,47</sup>. This non-monotonic relationship can be appreciated in studies of error augmentation. For example, during reach adaptation to a visuomotor rotation, the rate of learning almost doubles when visual errors are amplified by a gain of 2<sup>48</sup>. However, when visual errors are amplified by a gain of 3, the rate of learning returns to the level associated with veridical visual feedback<sup>48</sup>. Given this non-monotonic relationship between adaptation and error magnitude, it is possible that muscle co-contraction increases the rate of adaptation by reducing errors to a size that induces greater learning.

Previous studies of error sensitivity<sup>16,34,36,44,46,47</sup> did not examine its dependence on muscle co-contraction. Therefore, it is possible that error sensitivity is a function of muscle co-contraction, such that as muscle co-contraction increases, single-trial adaptation is maximized by progressively smaller errors. For the sensorimotor system to implement such an error-sensitivity function, it would need to have knowledge of the effects of muscle co-contraction on the statistics of kinematic errors. Such knowledge has been postulated in normative models of impedance control, in which muscle co-contraction emerges as a strategy to minimize the expected cost of movement by reducing uncertainty in the forward dynamics model<sup>8</sup>. If error sensitivity were a function of muscle co-contraction, when kinematic errors are experienced, the sensorimotor system could appropriately assign credit between the forward dynamics model and sensorimotor noise, controlling for the level of muscle co-contraction. Interestingly, the forward dynamics model, which maps current states and motor commands to next states, is also a function of muscle co-contraction (a state determined by a motor command), and so it is an open question to what degree adaptation under increased co-contraction generalizes to other co-contraction levels.

All participants were able to form an internal model of the force field, as demonstrated by adaptation on channel trials and after-effects in the post-exposure phase. This is consistent with previous studies that have demonstrated that people can adapt to viscous force fields that are superimposed on background loads<sup>49,50</sup>. It has even been suggested that people partition force fields into dynamic and static components<sup>49</sup>, which could allow velocity-dependent dynamics to be learned across a range of muscle load states. However, not all muscle states appear to be available to the internal model. For example, after recovery from fatigue, muscles initially generate excessive force, as if the internal model is still producing motor commands appropriate for fatigued muscles<sup>51</sup>.

In our study, although participants made movements in four directions, we only measured adaptation for one direction to limit the length of the experiment. Given that endpoint stiffness is in general anisotropic<sup>10,52,53</sup>, adaptation may similarly have been anisotropic, giving rise to an adaptation ellipse. Moreover, the geometry (size, shape, orientation) of this adaptation ellipse, like the geometry of the stiffness ellipse<sup>10,52,53</sup>, may be modifiable, allowing the rate of adaptation to be modulated in a direction-dependent manner. For example, by orienting the major axis of the stiffness ellipse in the direction of greatest unpredictability in the environment, it may be possible to maximize the rate of adaptation in the direction it is needed most: where errors are largest.

It remains an open question whether muscle co-contraction can modulate adaptation independently of the errors experienced. Feedback error learning (FEL)<sup>31,54</sup> provides one possible mechanism by which this could occur. According to FEL, the feedback response to error acts as a teaching signal for motor adaptation. Consistent with FEL, single-trial adaptation has been shown to correlate with the magnitude of the feedback response<sup>14</sup>. Given that feedback gains scale with muscle activity<sup>55</sup> and feedback responses are potentiated when muscles are co-contracted<sup>56–58</sup>, adaptation to an error of a given magnitude should be larger when muscles are co-contracted.

In conclusion, we have shown that muscle co-contraction accelerates dynamic motor learning, despite reducing kinematic errors, which are the main driving force behind adaptation. Muscle co-contraction therefore improves both kinematic accuracy and dynamic learning, simultaneously enhancing present and future motor performance. The modifiable nature of muscle co-contraction suggests that the rate of motor adaptation can be actively modulated.

## Data Availability

Data that support the findings of this study are available from the corresponding author on request.

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## Author Contributions

All authors conceived and designed the experiments. J.H. performed the experiments. All authors wrote the paper, discussed the results and edited the manuscript.

## Additional Information

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# **Increasing muscle co-contraction speeds up internal model acquisition during dynamic motor learning**

## **Supplementary Information**

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## Instructions to participants

### *Instructions to the stiff group (provided prior to commencement of the pulse phase)*

I'm now going to describe the next phase of the experiment. Each trial will start when you place the green cursor in the center of the white home position. A brief force will then be applied to the handle of the robot in a randomly chosen direction. This will only last for half a second. Your task will be to keep the handle as still as possible. To keep the handle still, you will need to stiffen up your arm by tensing the muscles in your hand, arm and chest. If on one trial you get pushed outside of the red ring that surrounds the white home position, you should try to stiffen up more on the next trial to help you stay within the ring. Ideally, you should keep the handle as still and as close to the center of the white home position as possible.

Once you have completed this phase of the experiment, all subsequent trials will be similar to the ones you've just done; that is, they will involve you making straight reaching movements to one of four targets. However, at some point your movements will be unexpectedly thrown off course by forces generated by the robot. Your task will be to keep your movements as straight as possible. In particular, you should try to stay as close as possible to the red lines that lead from the home position to the targets. You may find that stiffening up your arm will help you to do this. If at any point you feel as though your arm is too sore or tired, just let me know and you can have a rest break immediately. You will automatically get a rest break every 5 minutes though and ideally you should wait until then in order to allow the experiment to run smoothly.

### *Instructions to the relaxed group (provided prior to commencement of the pulse phase)*

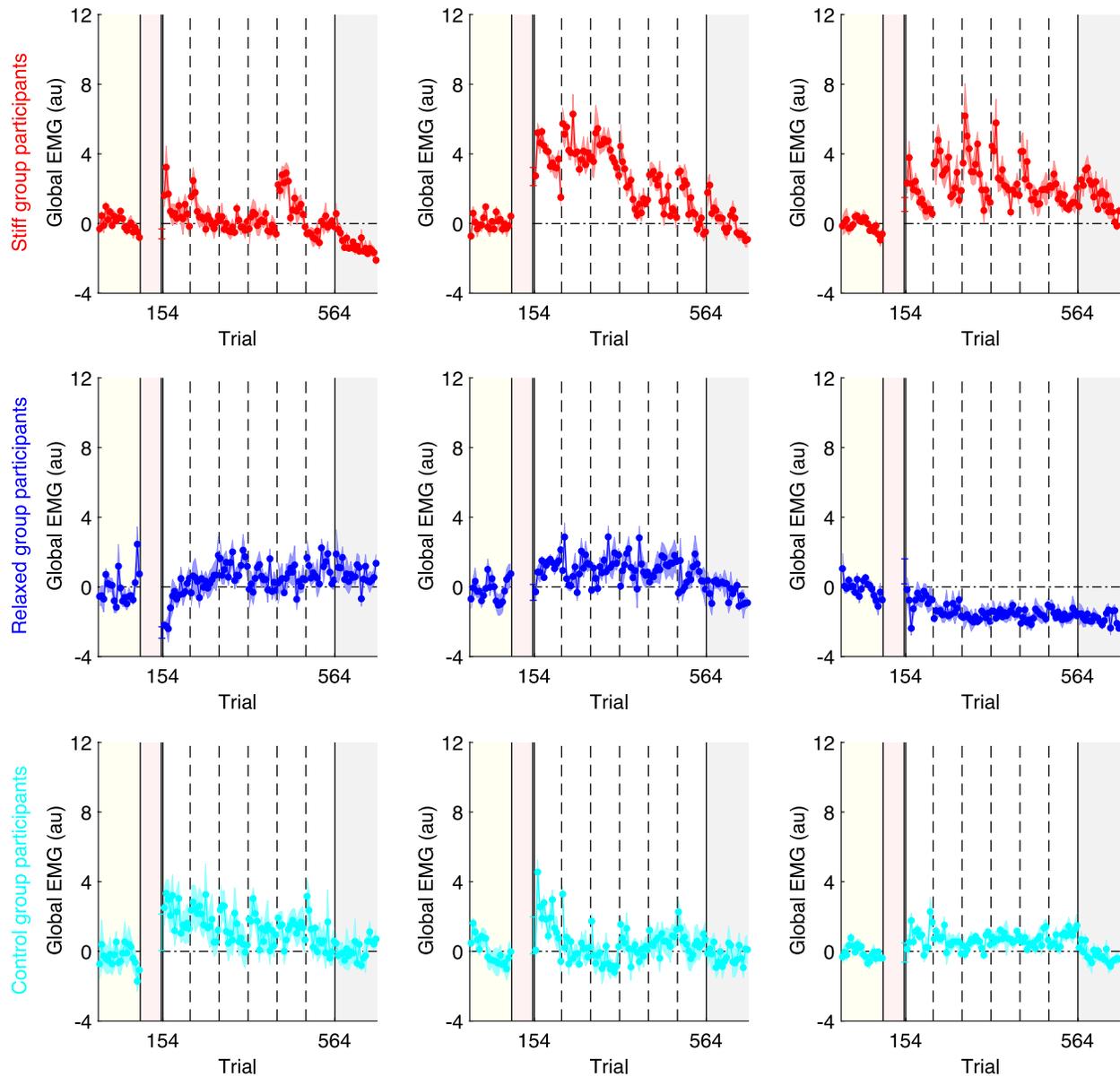
I'm now going to describe the next phase of the experiment. Each trial will start when you place the green cursor in the center of the white home position. A brief force will then be applied to the handle of the robot in a randomly chosen direction. This will only last for half a second. You should let your hand move freely to wherever the handle takes it. To do this, you should relax the muscles in your arm as much as possible so that you don't provide any resistance to the handle as it moves. Once the handle has stopped the robot will automatically bring you back to the central home position. Once again, you should keep your arm as relaxed as possible when this happens. Throughout all the trials of this phase you will see a red ring surrounding the white home position. This ring has no relevance and you should ignore it.

Once you have completed this phase of the experiment, all subsequent trials will be similar to the ones you've just done; that is, they will involve you making straight reaching movements to one of four targets. However, at some point your movements will be unexpectedly thrown off course by forces generated by the robot. Try not to stiffen up when this happens. If you relax your arm you will find that your movements will naturally become straighter as you adapt to the forces.

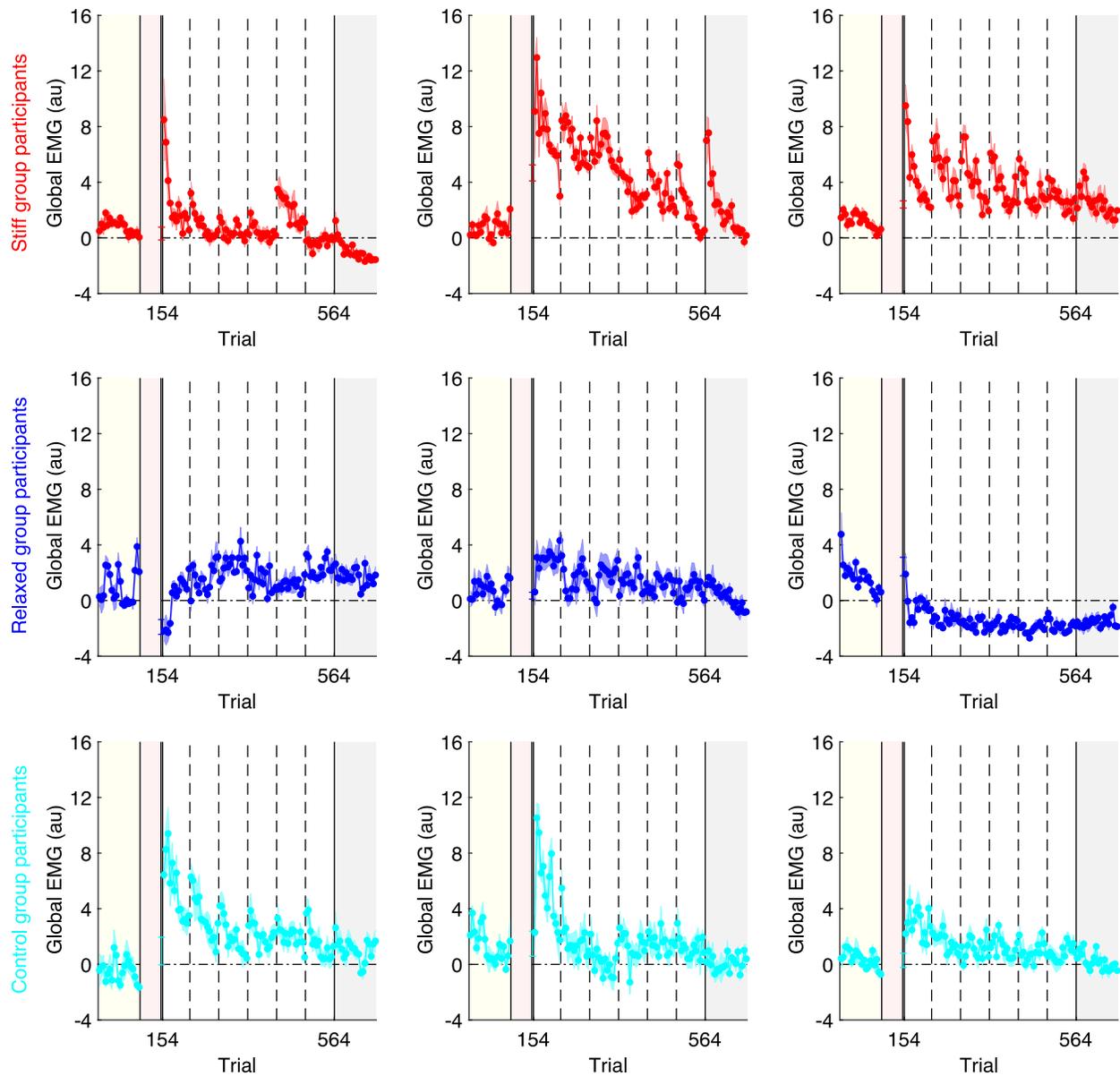
### *Instructions to the control group (provided prior to commencement of the experiment)*

At some point during the experiment, your movements will be unexpectedly thrown off course by forces generated by the robot. When this happens, keep trying to reach the target, and you will find that you will naturally adapt to the forces over time.

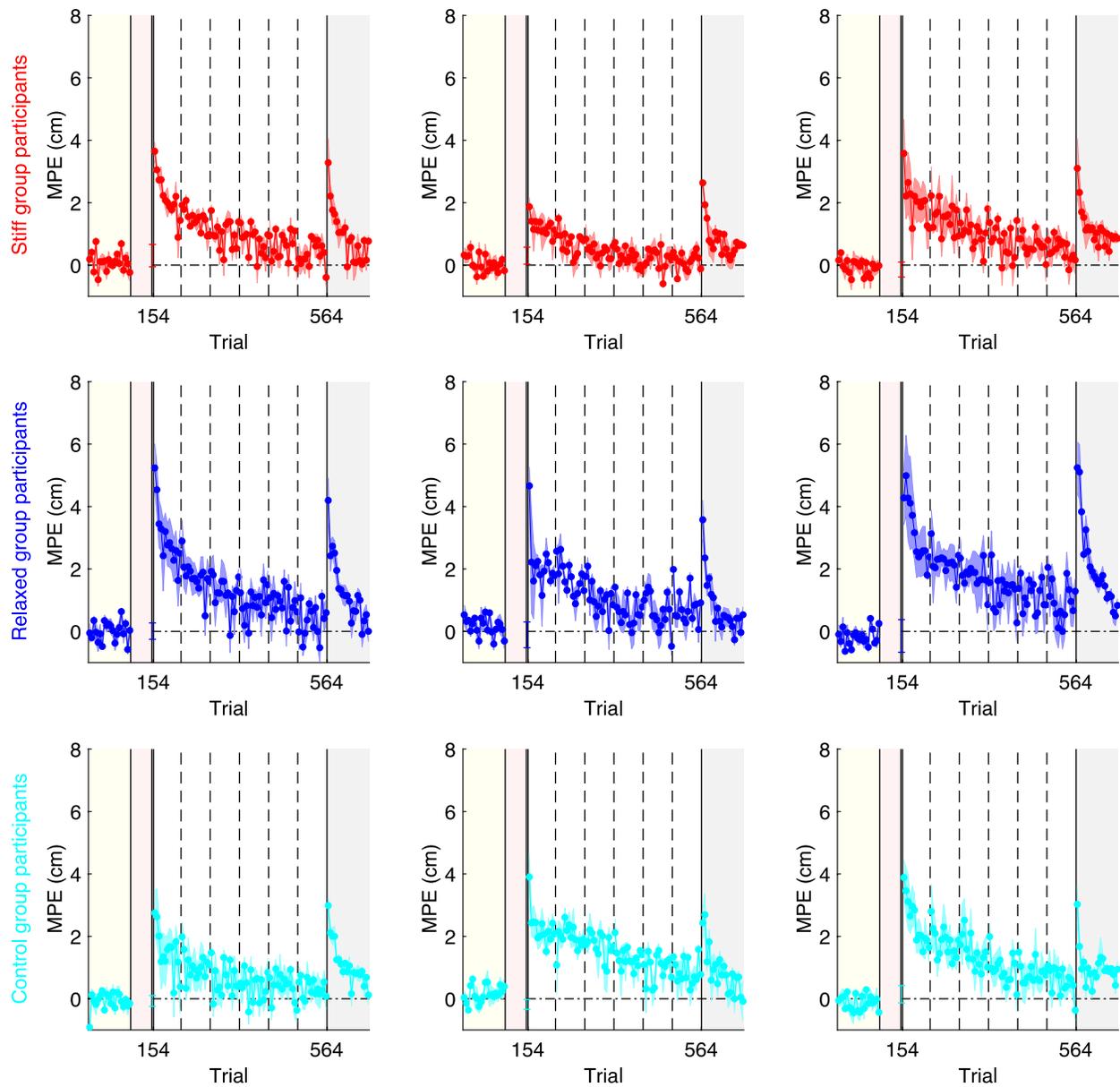
## Supplementary Figures



**Supplementary Figure 1 | Global EMG in the early movement period over the course of the experiment.** Global EMG was calculated on each trial from -200 ms to 130 ms relative to movement onset and plotted as mean  $\pm$  s.e.m over a block of five trials. Each subplot shows data from an individual participant in either the stiff (red), relaxed (blue) or control (cyan) groups (the same nine participants are shown in Supplementary Fig. 1-4). Shaded regions indicate the pre-exposure (yellow), pulse (red), exposure (white), and post-exposure (gray) phases. Vertical dashed lines denote rest breaks.

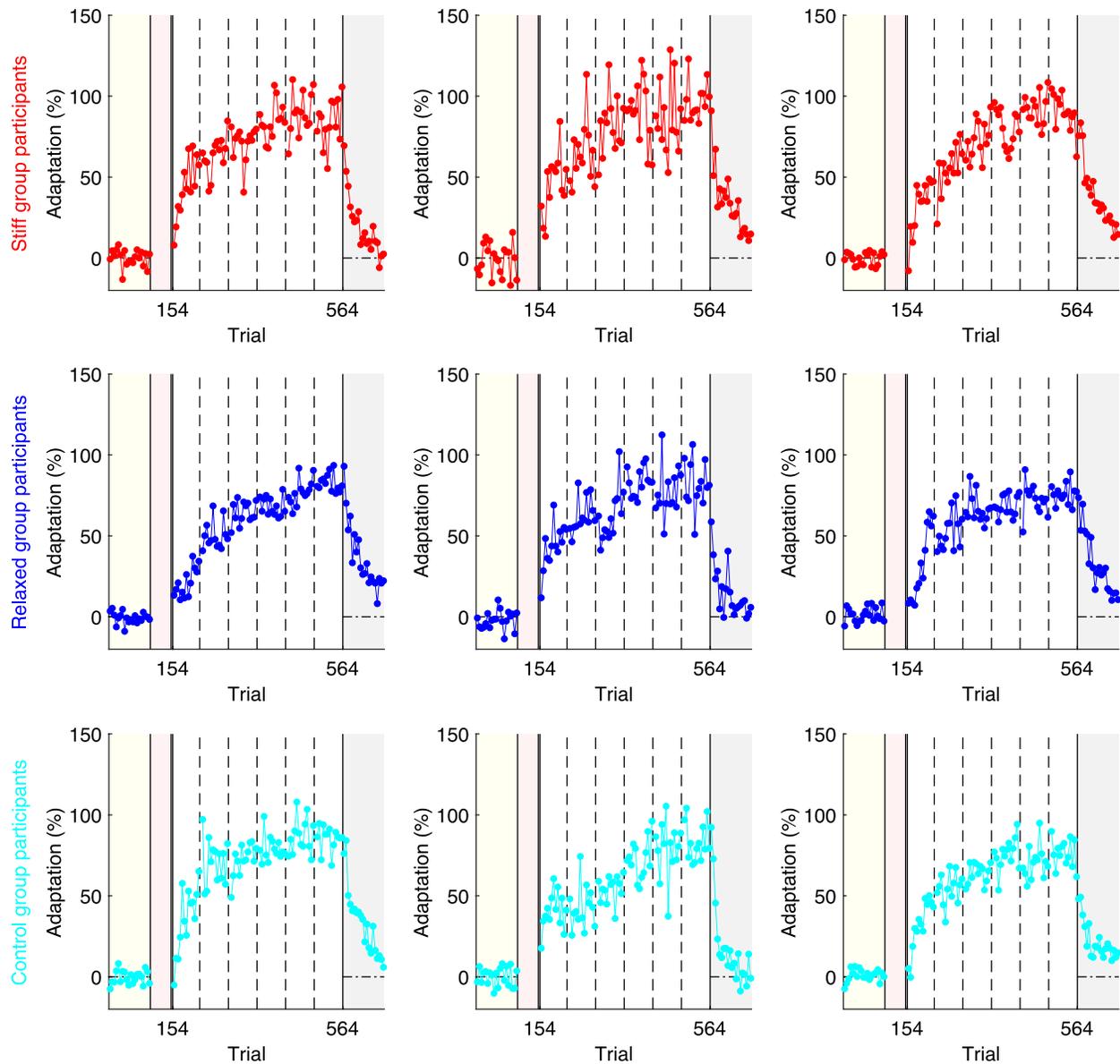


**Supplementary Figure 2 | Global EMG in the later movement period over the course of the experiment.** Global EMG was calculated on each trial from 130 ms to 400 ms relative to movement onset and plotted as mean  $\pm$  s.e.m over a block of five trials. Each subplot shows data from an individual participant in either the stiff (red), relaxed (blue) or control (cyan) groups (the same nine participants are shown in Supplementary Fig. 1-4). Shaded regions indicate the pre-exposure (yellow), pulse (red), exposure (white), and post-exposure (gray) phases. Vertical dashed lines denote rest breaks.



**Supplementary Figure 3 | Kinematic adaptation over the course of the experiment.**

Maximum perpendicular error (MPE) was calculated on each trial and plotted as mean  $\pm$  s.e.m over a block of five trials. Each subplot shows data from an individual participant in either the stiff (red), relaxed (blue) or control (cyan) groups (the same nine participants are shown in Supplementary Fig. 1-4). Shaded regions indicate the pre-exposure (yellow), pulse (red), exposure (white), and post-exposure (gray) phases. Vertical dashed lines denote rest breaks.



**Supplementary Figure 4 | Dynamic adaptation over the course of the experiment.**

Adaptation measured on channel trials. Each subplot shows data from an individual participant in either the stiff (red), relaxed (blue) or control (cyan) groups (the same nine participants are shown in Supplementary Fig. 1-4). Shaded regions indicate the pre-exposure (yellow), pulse (red), exposure (white), and post-exposure (gray) phases. Vertical dashed lines denote rest breaks.